Abstract
Crizotinib is an orally available tyrosine kinase inhibitor, approved for treatment of anaplastic lymphoma kinase (ALK) and c-ros oncogene 1 (ROS1) rearrangement-positive non-small cell lung cancer (NSCLC). According to the product leaflet, crizotinib capsules should be swallowed whole, and should not be crushed, dissolved or opened. However, this manner of administration is not always possible. At present, literature is lacking regarding the absorption of crizotinib via percutaneous endoscopic jejunostomy (PEJ) tube. We report a case of a patient with ALK+ NSCLC who was administered crizotinib via PEJ tube. An adequate steady state crizotinib trough concentration was reached, resulting in a metabolic response. Safety for the caregiver was ensured since the administration of crizotinib was made without crushing or opening the capsule. This case supports the option for providing crizotinib via PEJ tube in patients who have ALK+ NSCLC and are unable to swallow whole capsules. This option might also apply to the administration of other ALK inhibitors.

Keywords: Crizotinib, ALK inhibitor, percutaneous endoscopic jejunostomy tube, pharmacokinetics, non-small cell lung cancer.

To our knowledge, this is the first report of a patient having achieved an adequate steady state crizotinib trough concentration after receiving crizotinib administered via PEJ tube.

**Case Presentation**

A 36-year-old female patient, known with ALK+ translocated NSCLC, presented in 2016 with recurrent disease with distant metastasis after previous chemoradiation and resection. The preferred treatment was crizotinib. However, due to complications of the previous treatment, oral intake was not possible and she received nutrition by PEJ. Crizotinib was started at a dose of 250 mg twice daily, which was administered via PEJ tube as a dispersion in lukewarm water. The method of administration was similar as being described by Tamai *et al* [2]. Since the specific site of absorption of crizotinib within the gastrointestinal system is currently unknown, it was decided to determine a steady state crizotinib trough concentration at day 20 of therapy [1]. The crizotinib trough concentration was analyzed by a validated high-performance liquid chromatography-tandem mass spectrometry (HPLC-MS/MS) method in a laboratory of the Netherlands Cancer Institute-Antoni van Leeuwenhoek (NKI-AVL). The crizotinib trough concentration reached was 304 ng/mL. Therapy was continued at a dose of 250 mg twice daily. No tube obstruction or adverse events were seen. Tumour response was radiographic assessed by positron emission tomography-computed tomography (PET-CT) according to RECIST version 1.0 and revealed a metabolic response after two months of treatment.

**Discussion**

The steady state crizotinib trough concentration reached in this patient, 304 ng/mL, was in line with crizotinib concentrations described in literature, ranging from respectively 244 to 848 ng/mL in Asian populations [3] and 242 to 319 ng/mL in Caucasian populations [4].

We had access to the electronic patient record and the pharmacy dispensing data which contained large amount of information regarding multiple factors known to influence the exposure of crizotinib. First of all, we performed a medication review since crizotinib is known to be a CYP3A4 substrate, a moderate time-dependent inhibitor, and a weak inducer [5]. We found no drug-drug interactions. However, our patient was using pantoprazole 40 mg once daily, which was administered via PEJ tube [1,6]. The jejunal pH is significantly higher than the gastric pH and the aqueous solubility of crizotinib is pH dependent, with low (acidic) pH resulting in higher solubility [1,7]. However, the exposure of crizotinib was not affected in a significant manner by the use of pantoprazole and the administration via jejunal tube. This result is in line with literature [1,6]. Furthermore, since our patient was fed by enteral nutrition, the nutritional composition over time was stable. Although the effect of food on steady state crizotinib exposure is currently unknown, it is described that a high-fat meal decreases the systemic exposure by an average of 15% compared to fasting conditions [1]. Moreover, the patient did not use any food (e.g. grapefruit juice) or herbal supplements (e.g. St. John’s wort). The pharmacokinetic covariates of the patient (Caucasian female, 36 years, ±55 kg, non-smoker, normal renal and hepatic function) were not expected to significantly influence the exposure of crizotinib [8].

A multitude of factors need to be accounted for when planning to deliver drugs by feeding tubes. Product alterations necessary to allow tube delivery may interfere with product stability, compatibility with concomitant medications and tolerability, as well as pharmacokinetic parameters [9]. However, crizotinib is an immediate-release formulated capsule and can be administered by any tube comprised of polyurethane, polyvinyl chloride (PVC) or silicone [1,10]. The crizotinib administration was performed via dissolution of the formulated capsule into a 20 mL syringe containing lukewarm water [10]. No specific safety precautions were necessary since the capsule did not have to be crushed or opened [10]. However, it is important for caregivers, who handle the medication, to avoid skin contact with crizotinib. Wearing disposable gloves should be advised in order to safely transfer the capsule from the package to the syringe. It is possible to assume that the described administration method can also be applied for second and third-generation ALK inhibitors, candidates being alectinib, brigatinib, ceritinib and...
lorlatinib [11]. Similar to other tyrosine kinase inhibitors, these agents have been developed as immediate-release formulated capsules and tablets, and not as delayed-, extended-, or sustained-release formulations [12-14]. In addition, all agents are not enteric coated. The instruction to take capsules and tablets without crushing, dissolving or opening seems to be provided to minimize environmental exposure of these agents. Adequate absorption cannot be guaranteed by administrating these agents as dispersions via NG, PEG or PEJ enteric feeding tubes. Similar to our report, concentration measurements during steady state may be performed to determine exposure. However, this is only possible when validated analytical methods are available. Future pharmacokinetic studies are warranted to investigate these methods of administration.

Conclusion
In this case report, we demonstrate adequate crizotinib absorption after administration via PEJ tube with preservation of the steady state trough crizotinib concentration. This method of administration might be applicable to other ALK inhibitors.

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Conflicts of Interest
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